

Contents lists available at ScienceDirect

International Dairy Journal

journal homepage: www.elsevier.com/locate/idairyj



Review

Nutritional modulation of gut microbiota in the context of obesity and insulin resistance: Potential interest of prebiotics

Nathalie M. Delzenne*, Patrice D. Cani

Université catholique de Louvain, Louvain Drug Research Institute, Metabolism and Nutrition Research Group, PMNT Unit-7369, Av. E. Mounier, 73/69, B-1200 Brussels, Belgium

ABSTRACT

Obesity in humans leads to changes in the composition of gut microbiota, some of those changes being reversed upon dieting and changes in dietary habits. The studies devoted to understand how gut microbes control host energy homeostasis are of interest, in order to estimate how specific nutrients that induce changes in gut microbiota composition and/or activity – such as prebiotics – could be relevant in the management of obesity and related disorders. This review presents the potential molecular mechanisms allowing the gut microbiota to control host energy homeostasis, and presents the potential mechanisms evoked in the improvement of obesity by colonic nutrients that target the gut microbiota. It also discusses the relevance of this new area of research in human nutrition and health.

© 2009 Elsevier Ltd. All rights reserved.

Contents

)
1.	Changes in gut microbiota composition occur upon obesity and related metabolic disorders	.,277
2.	Gut microbiota participates to the regulation of host energy homeostasis	278
3.	Food components that modulate the gut microbiota have an impact on obesity and related disorders; the potential interest of prebiotics	278
4.	Probiotics and prebiotics approach in the management of obesity and associated diseases in human	279
	Conclusion	
	References	270

1. Changes in gut microbiota composition occur upon obesity and related metabolic disorders

Obesity is associated with a cluster of metabolic disorders including glucose intolerance, insulin resistance, type 2 diabetes, hypertension, dyslipidemia, fibrinolysis disorders, epithelial dysfunction, atherosclerosis, cardiovascular diseases, non-alcoholic fatty liver diseases (NAFLD) and non-alcoholic steatohepatitis (NASH) (Eckel, Grundy, & Zimmet, 2005; Ogden, Yanovski, Carroll, & Flegal, 2007). The adverse health consequences of weight gain and obesity are especially prominent following prolonged periods of positive energy balance and is mostly associated with a high-fat diet ingestion in our Western countries.

Results obtained both in rodents and humans, suggest that obesity is associated with an altered composition of gut microbiota

(for review see Ley, 2009). Obesity is characterized by a reduced level of gut microbes phylogenic diversity (Turnbaugh et al., 2009), and by a division wide shifts of the two major phyla: more Firmicutes characterize obese versus lean individuals; some authors show a drop in Bacteroidetes upon obesity, whereas other authors show no change or even an increase of Bacteroidetes in overweight (Collado, Isolauri, Laitinen, & Salminen, 2008; Duncan et al., 2008; Ley, Turnbaugh, Klein, & Gordon, 2006; Turnbaugh et al., 2006). The level of fecal Bifidobacterium spp. was shown to be higher in children remaining normal weight at the age of seven, whereas it was not the case in overweight children. In addition, the authors observed that the Staphylococcus aureus count was lower in children who maintain a normal weight than in overweight (Kalliomaki, Collado, Salminen, & Isolauri, 2008). For that reason, it has been proposed that changes in specific bacteria could play a role in the development of obesity.

A question remains open: are the changes in gut microbiota in overweight or obese individuals due to the obesity per se, or due to changes in nutritional habits in obese people?

^{*} Corresponding author. Tel.: +32 2 764 73 97; fax: +32 2 764 73 59. E-mail address: Nathalie.Delzenne@uclouvain.be (N.M. Delzenne).

Interestingly, the first studies have shown that dieting – whatever are the nutritional changes (low carbohydrates or low fat diet) – was able per se to reverse the shift in Firmicutes/Bacteroidetes (Ley et al., 2006). Other studies have correlated changes in gut microbiota with carbohydrate intake (for review see Ley, 2009). Data obtained in animals suggest that a high-fat diet – and not the obese state per se – can modulate microbiota composition towards an increase in Firmicutes and a proportional decrease in Bacteroidetes (Hilderbrandt, Hoffman, & Sherrill-Mix, 2009)

2. Gut microbiota participates to the regulation of host energy homeostasis

Evidence on the role of the gut microbiota on energy harvesting from the diet came from studies performed in germ-free mice (Bäckhed et al., 2004; Ley, 2009). Bäckhed et al. (2004) found that conventionally raised mice contained 40% more total body fat and 47% higher gonadal fat content than germ-free mice. Several pathways are proposed to explain that the presence of the gut microbiota drives the increase in fat mass - shown both in animals fed a standard carbohydrate-rich or a fat-rich diet (Cani & Delzenne, 2009a, 2009b; Ley, 2009; Musso, Gambino, & Cassader, 2009). The gut microbiota might promote intestinal monosaccharide absorption, energy extraction from non-digestible food components (via short chain fatty acids (SCFA) production through the fermentation), hepatic de novo lipogenesis, and adipocyte fatty acid storage; this latter effect is driven through lipoprotein lipase regulation (LPL) via the suppression of intestinal expression of an LPL inhibitor (FIAF, fasting-induced adipose factor). The second pathway, further explored the underlying mechanisms related to the fact that germfree mice are protected against high-fat diet-induced obesity and associated metabolic disorders, independent on energy harvesting (Bäckhed, Manchester, Semenkovich, & Gordon, 2007). The authors have shown that proposed form their data that the presence of the gut microbiota suppresses the liver and skeletal muscle AMP-activated protein kinase (AMPK) -dependent fatty acid oxidation, thereby lessening an important catabolic pathway. A third pathway, involving the gut microbiota fermentation end-products, namely the short chain fatty acids (SCFA), has been recently proposed. SCFA act not only as energy substrates for the host, but also as signalling molecules. They are ligands for at least two G protein-coupled receptors, GPR41 and GPR43. Samuel et al. (2008) have demonstrated that $GPR41^{-/-}$ mice colonized with a model of fermentative microbial community (Bacteroides thetaiotaomicron and Methanobrevibacter smithii) did not gain fat mass at the same extent as wildtype littermates did. The authors proposed that in the absence of GPR41 signalling, the reduced plasma PYY levels promotes gut motility and reduces energy harvest from the diet. However, this last hypothesis - based on the fact that modulation of PYY level influences intestinal transit, is not in accordance with the fact that the modulation of gut microbiota by prebiotics (discussed later in this paper) increased SCFA concentration in the caecum but also increased plasma PYY levels, a mechanism probably contributing to the reduction of food intake and fat mass development upon prebiotic treatment (Cani & Delzenne, 2009a; Delzenne, Cani, & Nevrinck, 2007).

Metabolic diseases are commonly linked to inflammatory processes and the gut microbiota can also play a role in this process. In fact, animal and human data indicate that an increased level of circulating lipopolysaccharides (LPS) – called endotoxemia – is observed upon high-fat feeding, and is linked to disturbed glucose homeostasis (Cani, et al., 2007a). The first study examining the kinetics of baseline endotoxemia concentrations in healthy human subjects has been published by Erridge, Attina, Spickett, and Webb (2007). In this study, the authors found that a high-fat meal induces

a metabolic endotoxemia leading to LPS concentrations that may be sufficient to induce some degree of cellular activation of monocytes in vitro. In addition, endotoxemia correlates with fasting insulin in non-diabetes patients, and is two-fold higher in type 2 diabetes patients group than in the non-diabetic subjects (Creely et al., 2007). A positive correlation has also been demonstrated between plasma endotoxin levels and energy/or fat intake in humans (Amar et al., 2008).

Experimental data previously suggested that LPS, derived from gram negative bacteria present in the gut, plays a key role in driving fat mass development, insulin resistance and systemic inflammation, since all those metabolic alterations do no occur in CD14receptor knock out animals (CD14 being a key receptor component allowing LPS cellular response) (Cani, et al., 2007a), and are avoided in obese animals treated with large spectrum antibiotics, a treatment blunting intestinal LPS content (Cani et al., 2008). LPS is transported from the intestine towards target tissues by a mechanism facilitated by lipoproteins, notably chylomicrons freshly synthesized from epithelial intestinal cells in response to fat feeding (Ghoshal et al., 2009). Other mechanisms are also involved in the increased endotoxemia upon obesity, such as an increase in gut permeability linked to the decrease in expression and of the adequate repartition of tight junction proteins (Zonula occludens 1 and Occludin) (Cani et al., 2008; Cani et al., 2009b).

The link between gut microbiota, LPS translocation upon obesity and the development of adiposity could involve serum amyloid (SAA) protein. The level of this protein is increased in obese individuals and in mice fed a high-fat diet. Results comparing conventionally raised and germ-free mice have shown that in the presence of gut microbes, SAA3 is elevated in the adipose tissue of mice, and this increase requires an LPS-mediated mechanism (Reigstad, Lundén, Felin, & Bäckhed, 2009).

Form this section, one could think that, finally, the presence of the gut microbiota could be "harmful" when considering obesity and related disorders, as illustrated by the numerous processes by which, globally, gut microbes drive energy sparing and fat mass development. However, most of these data have been obtained through the comparison of germ-free versus conventional mice, two "extreme" situations, which cannot be reproduced through a nutritional approach. Moreover, as shown before, qualitative changes of the gut microbiota occur upon obesity, and it is well known that specific genders, species, or even strains of bacteria can exert drastically different effects related to host physiology. Therefore, we will show in the next sections, how "targeted" modification of gut microbiota is able to counteract obesity and associated disorders

3. Food components that modulate the gut microbiota have an impact on obesity and related disorders: the potential interest of prebiotics

In a series of experiments in mice fed a high-fat/carbohydrate free diet, we showed that such a dietary manipulation, leading to obesity and diabetes, changes bacterial populations in the intestinal microbiota, with a strong reduction in *Bifidobacterium* spp. numbers, a reduced *Bacteroides*-related bacteria, *Eubacterium* rectale-Clostridium coccoides group content (Cani, et al., 2007a, 2007c). In those studies, we found that among the different gut bacteria analyzed, metabolic endotoxemia correlated negatively with the bifidobacteria count (Cani, et al., 2007c). The administration of *Bifidobacterium* spp. as a probiotic has been shown to reduce the intestinal endotoxin levels and to improve mucosal barrier function (Griffiths et al., 2004; Wang et al., 2006, 2004). Another possibility to selectively modulate the gut microbiota is the prebiotic approach. The prebiotic concept refers to non-digestible dietary

ingredients that are selectively fermented by certain bacteria in the gastro-intestinal tract, thereby modifying the composition and/or activity of the gut microbiota with beneficial effects for host health (Gibson & Roberfroid, 1995; Roberfroid, 2007; Saulnier, Spinler, Gibson, & Versalovic, 2009). Prebiotics can be given orally (Tuohy, Rouzaud, Bruck, & Gibson, 2005) to specifically increase the gut bifidobacteria content of high-fat diet treated mice. Some prebiotics, such as fructans are also able to promote other interesting bacteria such as Faecalibacterium prausnitzii, for example (Ramirez-Farias et al., 2009), Since the first modulating effect on gut microbiota was established with bifidobacteria, the relation with the health effects has mostly been associated with the changes in this genus. We found that in prebiotic treated mice, Bifidobacterium spp. significantly and positively correlated with improved glucose homeostasis markers and normalized inflammatory tone (decreased metabolic endotoxemia, plasma and adipose tissue proinflammatory cytokines) (Cani, et al., 2007c).

Several mechanisms have been proposed in order to explain the positive effect of the prebiotic approach in obesity models. Among those hypotheses, the modulation of endocrine function occurs in prebiotic-fed animals is a phenomenon that contributes to the improvement of obesity and associated disorders (Cani & Delzenne, 2009a, 2009b; Cani, et al., 2006b). Endocrine L cells are present all along the gastro-intestinal tract. The expression of proglucagon gene in those cells leads to the secretion of different peptides, including glucagon-like peptides (GLP) 1 and 2, which play a role in host gut function and physiology. The ingestion of prebiotics has been shown to increase the number of L cells in the proximal colon of rats (Cani, Hoste, Guiot, & Delzenne, 2007b). The resulting increase in portal GLP-1 allows this peptide to regulate food intake and glucose homeostasis (Cani, et al., 2006b; Maurer, Chen, McPherson, & Reimer, 2009; Urias-Silvas et al., 2008). On the other hand, the increase in GLP-2 contributes to the improvement of gut barrier function and to the decrease in endotoxemia in obese mice (Cani, et al., 2009b).

The relevance of gut peptide modulation by prebiotics in humans is only poorly studied until now, but data obtained in humans have recently linked the increase in post-prandial gut peptide serum levels and the increase in satiety (Cani, et al., 2009a) and a decrease in fat mass development (Parnell & Reimer, 2009).

In addition to the role of LPS in the development of obesity, portal endotoxemia has been suggested to be a major risk in inducing hepatic inflammation in alcoholic liver diseases and NAFLD (Brun et al., 2007; Cani & Delzenne, 2009a, 2009b). Targeted modulation of gut microbiota by prebiotics can also contribute to the regulation of hepatic disorders associated with obesity. The use of prebiotics significantly reduces the hepatic triglyceride accumulation (steatosis) but also modifies hepatic inflammatory processes in the liver of different animal models of obesity (for review, Delzenne, Cani, & Neyrinck, 2008). Ma, Hua, and Li (2008) have pointed out a mechanism that could implicate an increase in specific hepatic immune cells that play a role in the maintenance of the equilibrium between pro-and anti-inflammatory cytokine production (NKT cells) in the probiotics-related improvement of glucose and lipid metabolism, but also hepatic disorders induced by a high-fat diet in mice. Similarly, the prebiotic approach is able to increase the phagocytic activity of hepatic fixed macrophages (Kupffer cells), thereby counteracting LPS-induced hepatotoxicity (Neyrinck, Alexiou, & Delzenne, 2004). The modulation of hepatic immune cell responsiveness by prebiotics in obese animals remains to be clarified.

Concerning human studies, only one paper has reported an improvement of hepatic alterations upon prebiotic treatment in non-alcoholic steatohepatitis patients (Daubioul, Horsmans, Lambert, Danse, & Delzenne, 2005).

4. Probiotics and prebiotics approach in the management of obesity and associated diseases in human

Even if the amount of relevant intervention studies remain scare in this field, namely due to the novelty of this concept, there are some data supporting the fact that both probiotic and prebiotic approaches could be interesting in the management of metabolic diseases associated with obesity. Interestingly, several physiological effects previously described in animals are also true when assessed upon mid term treatment with prebiotics in humans. A decrease in appetite and an increase in satiety, leading to a decrease in total energy intake, as well as a decrease in hepatic de novo lipogenesis. have been demonstrated in human volunteers fed with inulin-type prebiotics (16 g per day for several weeks) as compared to maltodextrin as a placebo (Cani, Joly, Horsmans, & Delzenne, 2006; Cani, Lecourt, et al., 2009; Letexier, Diraison, & Beylot, 2003). Prebiotics are however unable to drive an acute (48 h) change in food intake and appetite, thereby suggesting that the adaptation of the gut microbiota is required to have a physiological relevance in the control of food intake. A decrease in body mass index, linked to a modulation of gut peptides and appetite, have been shown upon long term treatment of overweight and obese patients (Abrams, Griffin, Hawthorne, & Ellis, 2007; Parnell & Reimer, 2009). An improvement of hepatic alterations has been reported in NASH patients receiving inulin-prebiotics versus placebo (maltodextrin) during 4-8 weeks (Daubioul et al., 2005).

5. Conclusion

The presence of saccharolytic gut bacteria in the gastro-intestinal tract is generally described to provide energy for the host, namely through the fermentation of the non-digestible carbohydrate and by promoting nutrients storage. However, targeted changes in the gut microbiota provided through the prebiotic approach can improve several metabolic disturbances occurring upon obesity. Most of the data obtained to date have been obtained in experimental animal studies, but promising effects are also shown in humans, thereby supporting the interest in the nutritional modulation of the gut microbiota in the management of metabolic diseases in overweight or obese patients. This concept could be applied to the prebiotic, but also to the probiotic approach; moreover, the metabolomic analysis will allow to select the potential new microbial targets related to obesity and related disorders in the future.

References

- Abrams, S. A., Griffin, I. J., Hawthorne, K. M., & Ellis, K. J. (2007). Effect of prebiotic supplementation and calcium intake on body mass index. *Journal of Pediatrics*, 151, 293–298.
- Amar, J., Burcelin, R., Ruidavets, J. B., Cani, P. D., Fauvel, J., Alessi, M. C., et al. (2008). Energy intake is associated with endotoxemia in apparently healthy men. American Journal of Clinical Nutrition, 87, 1219-1223.
- Bäckhed, F., Ding, H., Wang, T., Hooper, L. V., Koh, G. Y., Nagy, A., et al. (2004). The gut microbiota as an environmental factor that regulates fat storage, Proceedings of the National Academy of Sciences of the United States of America, 101, 15718–15723.
- Bäckhed, F., Manchester, J. K., Semenkovich, C. F., & Gordon, J. I. (2007). Mechanisms underlying the resistance to diet-induced obesity in germ free mice. Proceedings of the National Academy of Sciences of the United States of America, 104, 979–984.
- Brun, P., Castagliuolo, L., Leo, V. D., Buda, A., Pinzani, M., Palu, G., et al. (2007). Increased intestinal permeability in obese mice: new evidence in the pathogenesis of nonalcoholic steatohepatitis. *American Journal of Physiology-Gastro-intestinal and Liver Physiology*, 292, G518-G525.
- intestinal and Liver Physiology, 292, G518-G525.

 Cani, P. D., Amar, J., Iglesias, M. A., Poggi, M., Knauf, C., Bastelica, D., et al. (2007a).

 Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*, 56, 1761-1772.
- Cani, P. D., Bibiloni, R., Knauf, C., Waget, A., Neyrinck, A. M., Delzenne, N. M., et al. (2008). Changes in gut microbiota control metabolic endotoxemia-induced

- inflammation in high-fat diet-induced obesity and diabetes in mice. Diabetes, 57.1476-1481
- Cani, P. D., & Delzenne, N. M. (2009a). The role of the gut microbiota in energy metabolism and metabolic disease. Current Pharmaceutical Design, 15, 1546– 1558.
- Cani, P. D., & Delzenne, N. M. (2009b). Interplay between obesity and associated metabolic disorders: new insights into the gut microbiota. Current Opinion in Pharmacology, 9, 737-743.
- Cani, P. D., Hoste, S., Guiot, Y., & Delzenne, N. M. (2007b). Dietary non-digestible carbohydrates promote L-cell differentiation in the proximal colon of rats. British Journal of Nutrition, 98, 32–37.
 Cani, P. D., Joly, E., Horsmans, Y., & Delzenne, N. M. (2006a). Oligofructose promotes
- Cani, P. D., Joly, E., Horsmans, Y., & Delzenne, N. M. (2006a). Oligofructose promotes satiety in healthy human: a pilot study. European Journal of Clinical Nutrition, 60, 567-572
- Cani, P. D., Knauf, C., Iglesias, M. A., Drucker, D. J., Delzenne, N. M., & Burcelin, R. (2006b). Improvement of glucose tolerance and hepatic insulin sensitivity by oligofructose requires a functional glucagon-like peptide 1 receptor. *Diabetes*, 55, 1484-1490.
- Cani, P. D., Lecourt, E., Dewulf, E. M., Sohet, F. M., Pachikian, B. D., Naslain, D., et al. (2009a). Gut microbiota fermentation of prebiotics increases satietogenic and incretin gut peptide production with consequences for appetite sensation and glucose response after a meal. American Journal of Clinical Nutrition, 90, 1236– 1243.
- Cani, P. D., Neyrinck, A. M., Fava, F., Knauf, C., Burcelin, R. G., Tuohy, K. M., et al. (2007c). Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. Diabetologia, 50, 2374-2383.
- Cani, P. D., Possemiers, S., Van de, W. T., Guiot, Y., Everard, A., Rottier, O., et al. (2009b). Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. Gut. 58, 1091-1103.
- Collado, M. C., Isolauri, E., Laitinen, K., & Salminen, S. (2008). Distinct composition of gut microbiota during pregnancy in overweight and normal-weight women. American Journal of Clinical Nutrition, 88, 894-899.
- Creely, S. J., McTernan, P. G., Kusminski, C. M., Fisher, M., da Silva, N. F., Khanolkar, M., et al. (2007). Lipopolysaccharide activates an innate immune system response in human adipose tissue in obesity and type 2 diabetes. American Journal of Physiology – Endocrinology and Metabolism, 292, E740–E747.
- Daubioul, C. A., Horsmans, Y., Lambett, P., Danse, E., & Delzenne, N. M. (2005). Effects of oligofructose on glucose and lipid metabolism in patients with nonalcoholic steatohepatitis: results of a pilot study. European Journal of Clinical Nutrition. 59, 723-726.
- Delzenne, N., Cani, P. D., & Neyrinck, A. (2007). Gut peptides and energy metabolism: experimental data with inulin and oligofructose. Journal of Nutrition, 137, 2547S-2551S.
- Delzenne, N., Cani, P. D., & Neyrinck, A. (2008). Prebiotics and lipid metabolism. In J. Versalovic, & M. Wilson (Eds.), Therapeutic microbiology: Probiotics and related strategies (pp. 183-192). Washington, DC: ASM Press.
- Duncan, S. H., Lobley, G. E., Holtrop, G., Ince, J., Johnstone, A. M., Louis, P., et al. (2008). Human colonic microbiota associated with diet, obesity and weight loss, International Journal of Obesity (Jondon), 32, 1720-1724.
- International Journal of Obesity (London), 32, 1720-1724.

 Eckel, R. H., Grundy, S. M., & Zimmet, P. Z. (2005). The metabolic syndrome. Lancet, 365, 1415-1428.
- Erridge, C., Attina, T., Spickett, C. M., & Webb, D. J. (2007). A high-fat meal induces low-grade endotoxemia: evidence of a novel mechanism of postprandial inflammation. American Journal of Clinical Nutrition, 86, 1286-1292.
- Ghoshal, S., Witta, J., Zhong, J., de Villiers, W., & Eckhardt, E. (2009). Chylomicrons promote intestinal absorption of lipopolysaccharides. *Journal of Lipid Research*, 50, 90-97.
- Gibson, G. R., & Roberfroid, M. B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *Journal of Nutrition*, 125, 1401–1412.
- Griffiths, E. A., Duffy, L. C., Schanbacher, F. L., Qiao, H., Dryja, D., Leavens, A., et al. (2004). In vivo effects of bifidobacteria and lactoferrin on gut endotoxin concentration and mucosal immunity in Balb/c mice. Digestive Diseases and Sciences, 49, 579-589.

- Hilderbrandt, M. A., Hoffman, C., & Sherrill-Mix, S. A. (2009). High fat fiet determines the composition of the murine gut microbiome independently of obesity. Gastroenterology. doi: 10.1052/j.gastro.2009.08.042.
- Gastroenterology. doi:10.1052/j.gastro.2009.08.042.
 Kalliomaki, M., Collado, M. C., Salminen, S., & Isolauri, E. (2008). Early differences in fecal microbiota composition in children may predict overweight. American Journal of Clinical Nutrition. 87. 534–538.
- Journal of Clinical Nutrition, 87, 534-538.

 Letexier, D., Diraison, F., & Beylot, M. (2003). Addition of inulin to a moderately high-carbohydrate diet reduces hepatic lipogenesis and plasma triacylglycerol concentrations in humans. American Journal of Clinical Nutrition, 77, 559-564.
- Ley, R. (2009). Obesity and the human microbiome. Current Opinion in Gastroenterology. doi:10.1097/MOG.0b013e328333d751.
- Ley, R. E., Turnbaugh, P. J., Klein, S., & Gordon, J. I. (2006). Microbial ecology: human gut microbes associated with obesity. Nature, 444, 1022–1023.
- Ma, X., Hua, J., & Li, Z. (2008). Probiotics improve high fat diet-induced hepatic steatosis and insulin resistance by increasing hepatic NKT cells. *Journal of Hepatology*, 49, 821–830.
- Maurer, A. D., Chen, Q., McPherson, C., & Reimer, R. A. (2009). Changes in satiety hormones and expression of genes involved in glucose and lipid metabolism in rats weaned onto diets high in fibre or protein reflect susceptibility to increased fat mass in adulthood. Journal of Physiology, 587, 679-691.
- Musso, G., Gambino, R., & Cassader, M. (2009). Gut microbiota as a regulator of energy homeostasis and ectopic fat deposition: mechanisms and implications for metabolic disorders. Current Opinion in Lipidology. doi:10.1097/MOL Ob013e3283347ebb.
- Neyrinck, A. M., Alexiou, H., & Delzenne, N. M. (2004). Kupffer cell activity is involved in the hepatoprotective effect of dietary oligofructose in rats with endotoxic shock. *Journal of Nutrition*, 134, 1124–1129.
- Ogden, C. L., Yanovski, S. Z., Carroll, M. D., & Flegal, K. M. (2007). The epidemiology
- of obesity. Gastroenterology, 132, 2087–2102.

 Parnell, J. A., & Reimer, R. A. (2009). Weight loss during oligofructose supplementation is associated with decreased ghrelin and increased peptide YY in overweight and obese adults. American Journal of Clinical Nutrition, 89, 1751–1759.

 Ramirez-Farias, C., Slezak, K., Fuller, Z., Duncan, A., Holtrop, G., & Louis, P. (2009).
- Ramirez-Farias, C., Slezak, K., Fuller, Z., Duncan, A., Holtrop, G., & Louis, P. (2009). Effect of inulin on the human gut microbiota: stimulation of Bifidobacterium adolescentis and Faecalibacterium prausnitzii. British Journal of Nutrition, 101, 541-550.
- Reigstad, C. S., Lundén, G. O., Felin, J., & Bäckhed, F. (2009). Regulation of serum amyloid A3 (SAA3) in mouse colonic epithelium and adipose tissue by the intestinal microbiota. PLoS One, 4, e5842.
- Roberfroid, M. (2007). Prebiotics: the concept revisited. *Journal of Nutrition*, 137, 830S-837S.
- Samuel, B. S., Shaito, A., Motoike, T., Rey, F. E., Backhed, F., Manchester, J. K., et al. (2008). Effects of the gut microbiota on host adiposity are modulated by the short chain fatty acid binding G protein-coupled receptor Gpr 41. Proceedings of the National Academy of Sciences United States of America, 105, 16767–16772.
- Saulnier, D. M., Spinler, J. K., Gibson, G. R., & Versalovic, J. (2009). Mechanisms of problosis and preblosis: considerations for enhanced functional foods. Current Opinion in Biotechnology, 20, 135-141.
- Tuohy, K. M., Rouzaud, G. C., Bruck, W. M., & Gibson, G. R. (2005). Modulation of the human gut microflora towards improved health using prebiotics assessment of efficacy. Current Pharmaceutical Design, 11, 75-90.
 Turnbaugh, P. J., Hamady, M., Yatsunenko, T., Cantarel, B. L., Duncan, A., Ley, R. E.,
- Turnbaugh, P. J., Hamady, M., Yatsunenko, T., Cantarel, B. L., Duncan, A., Ley, R. E., et al. (2009). A core gut microbiome in obese and lean twins. Nature, 457, 480–484.
- Turnbaugh, P. J., Ley, R. E., Mahowald, M. A., Magrini, V., Mardis, E. R., & Gordon, J. I. (2006). An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*, 444, 1027–1031.
 Urias-Silvas, J. E., Cani, P. D., Delmee, E., Neyrinck, A., Lopez, M. G., & Delzenne, N. M.
- Urias-Silvas, J. E., Cani, P. D., Delmee, E., Neyrinck, A., Lopez, M. G., & Delzenne, N. M. (2008). Physiological effects of dietary fructans extracted from Agave tequilana Gto, and Dasylirion spp. British Journal of Nutrition, 99, 254–261.
- Gto. and Dasylirion spp. British Journal of Nutrition, 99, 254–261.

 Wang, Z., Xiao, G., Yao, Y., Guo, S., Lu, K., & Sheng, Z. (2006). The role of bifidobacteria in gut barrier function after thermal injury in rats. Journal of Trauma, 61, 650–657.
- Wang, Z. T., Yao, Y. M., Xiao, G. X., & Sheng, Z. Y. (2004). Risk factors of development of gut-derived bacterial translocation in thermally injured rats. World Journal of Gastroenterology, 10, 1619–1624.