Use of Xanthinol Nicotinate as a co-treatment for radio- and chemo-therapy in experimental tumors

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The tumor micro-environment plays a key role in the tumor resistance to cytotoxic treatments. It has been demonstrated that it is possible to modulate the tumor microenvironment to potentiate anti-cancer therapy. Here, we made the hypothesis that the vasoactive agent xanthinol nicotinate (XN) could be an important modulator of the tumor perfusion and oxygenation. Using functional non invasive techniques (in vivo EPR oximetry and dynamic contrast enhanced MRI), we were able to define a time window in which tumor oxygenation, flow and permeability were significantly increased in the TLT tumor model implanted in muscles of mice. As a consequence of the alleviation of tumor hypoxia, we found out that XN was able to radiosensitize the tumors when applying 10 Gy of X-Rays during the reoxygenation of the tumors (enhancement in radiation response of L4). Moreover, the administration of cyclophosphamide (50 mg/kg) used as a chemotherapeutic agent was more efficient when applying the treatment after XN administration (enhancement in response to chemotherapy of 2.7). These results show the importance of the dynamic evolution of the tumor microenvironment on the response to treatments and that XN is an efficient modulator of the tumor hemodynamics that may potentiate cytotoxic treatments.

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It is now well established that regions within solid tumors experience mild to severe oxygen deprivation owing to aberrant vascular function. The hypoxic regions are associated with altered metabolism, as well as increased resistance to radiation and chemotherapy.1–3

New strategies for transiently opening the tumor vascular bed to alleviate tumor hypoxia, which is a source of resistance to radiotherapy via the 'oxygen effect', and improve the delivery of chemotherapeutic agents, are now considered in the pre-clinical setting. This so-called ‘pro-vascular’ approach has been named by opposition to the well known ‘anti-vascular’ and ‘anti-angiogenic’ strategies, using agents that are directed at the pre-existing vasculature suggesting the early effects of anti-angiogenic agents and pro-vascular agents might be similar.5–7 In the last years, our group has identified several pro-vascular agents that are able to improve the issue of radio and/or chemotherapy in experimental tumors, including NO-mediated agents, modulators of the thyroid status, and anti-angiogenic agents in their normalization phase, among others.6,8–13

In the current study, we hypothesized that xanthinol nicotinate (XN), a vasodilator used in the management of peripheral and cerebral vascular disorders, could improve tumor perfusion and oxygenation in order to get an improved radiotherapy and chemotherapy outcome. XN is the most potent form of niacin, the active form of vitamin B3, and acts as nicotinic acid. XN increases the cerebral metabolism of glucose, ATP levels and cerebral blood flow.14,15

We assessed the effect of XN on radiotherapy and cyclophosphamide chemotherapy in the TLT mouse tumor model. The first step was to monitor tumor pO2 after administration of XN by electron paramagnetic resonance (EPR) oximetry to determine the window of the vascular bed opening. As demonstrated by Patent Blue staining and dynamic contrast enhanced magnetic resonance imaging (DCE-MRI), the observed increase in pO2 was the signature of an increased tumor blood flow. These hemodynamic changes were then exploited in order to potentiate radiotherapy and chemotherapy, as demonstrated by the determination of regrowth delays.

Material and methods

Mice and tumors

Syngeneic TLT (Transplantable mouse Liver Tumor),16 were injected intramuscularly in the thigh of 5 week-old male NMRI mice (Animalerie facultaire, Université catholique de Louvain, Brussels). Tumor diameter was measured daily with a digital caliper. All animal experiments were performed in accordance with national animal care regulations.

XN and cyclophosphamide injections

XN was dissolved in saline and injected intraperitoneally at a final concentration of 75 mg/kg.13 The alkylating agent cyclophosphamide was dissolved in saline and injected intraperitoneally at the suboptimal dose of 50 mg/kg.7

Tumor oxygenation

Electronic Paramagnetic Resonance (EPR) Oximetry, using charcoal (CX0670-1, EM Science, Gibbstown, NJ) as the oxygen sensitive probe, was used to evaluate the tumor oxygenation.17 EPR oximetry relies on the oxygen-dependent broadening of the EPR line width of a paramagnetic oxygen sensor implanted in the tumor. This technique is designed for continuous measurement of the local pO2 without altering the local oxygen concentration, and allows repeated measurements from the same site over long periods of time. EPR spectra were recorded using an EPR spectrometer (Magnettech, Berlin, Germany) with a low frequency microwave bridge operating at 1.2 GHz and extended loop resonator. Mice were injected once in the center of the tumor 1 day before measurement using the suspension of charcoal (suspension of charcoal, 3% Arabic gum, 100 mg/ml, 50 μl injected, 1–25 μm particle size). The localized EPR measurements correspond to an average of pO2 values in a volume of ~10 mm3.17

Data acquisition was performed every 5 minutes for 1 hour after injection of XN (n = 5) or vehicle (n = 5). Tumor bearing mice were anesthetized using isoflurane (Fore`ne, Abott, Canada).
The perfusion was monitored in the time window during which intratumoral \( pO_2 \) was maximal, as determined by EPR oximetry after administration of XN and in the same time window after administration of vehicle. The perfusion was monitored via single-slice dynamic contrast-enhanced MRI at 4.7 T using the rapid-clearance blood pool agent P792 (Vistarem\textsuperscript{®}, Guerbet, Roissy, France).\textsuperscript{19} High resolution multi-slice \( T_2 \)-weighted spin echo anatomic imaging was performed just before dynamic contrast-enhanced imaging. Pixel-by-pixel values for \( K^{\text{trans}} \) (influx volume transfer constant, from plasma into the interstitial space, units of min\(^{-1}\)), \( V_p \) (blood plasma volume per unit volume of tissue, unitless), and \( K_{ep} \) (fractional rate of efflux from the interstitial space back to blood, units of min\(^{-1}\)) in tumor were calculated via tracer kinetic modeling of the dynamic contrast-enhanced data,\textsuperscript{20} and the resulting parametric maps for \( K^{\text{trans}} \), \( V_p \), and \( K_{ep} \) were generated. Statistical significance for \( V_p \) or \( K^{\text{trans}} \) identified “perfused” pixels, i.e., pixels to which the contrast agent P792 had access.\textsuperscript{6,19}

**Irradiation and tumor regrowth delay assay**

Irradiation was performed in the time window during which intratumoral \( pO_2 \) was maximal (i.e., 20 min after XN administration), as determined by EPR oximetry after administration of XN (group 1, \( n = 6 \)) and in the same time window after administration of vehicle (group 2, \( n = 5 \)). Non irradiated groups were also included (XN group, \( n = 5 \); and control group, \( n = 5 \)).

**Chemotherapy and tumor regrowth delay assay**

XN treated mice received a single dose of the alkylating agent cyclophosphamide (group 5, \( n = 5 \)) in the time window during which \( pO_2 \) and flow were maximal (i.e., 20 min after XN administration). Vehicle treated mice received the dose of alkylating agent during the same time window (group 6, \( n = 5 \)). Control experiments consisted in the injection of vehicle (group 7, \( n = 5 \)) and XN (group 3, \( n = 5 \)). After treatment, the tumor growth was determined daily by measuring tumor diameter until they reached a size of 16 mm, at which time the mice were sacrificed. A linear fit was performed between initial tumor size (7.5 ± 1 mm) and 16 mm which allowed determination of the time to reach a particular size for each mouse.

**Statistical analysis**

Results are given as means ± SEM values from \( n \) animals. Comparisons between groups were made with two-way ANOVA and a P value less than 0.05 was considered significant.

**Results**

**Effect of Xanthinol nicotinate on tumor \( pO_2 \)**

Intraperitoneal injection of XN rapidly and transiently modified tumor \( pO_2 \), as shown in Figure 1. Maximal \( pO_2 \) values were obtained 10 to 30 minutes after XN administration. These data determined the time window of vascular bed opening, which was used to design the “flow” experiments as well as as a rationale for scheduling the use of XN as a co-treatment in irradiation and cyclophosphamide chemotherapy experiments.

**Effect of Xanthinol nicotinate on tumor hemodynamics**

**Patent Blue staining.** A rough estimate of the tumor perfusion was carried out using the colored area observed in tumors after injection of a dye. Tumors with Xanthinol Nicotinate treatment stained more positive (\( n = 6 \); 65.5 ± 8.6% ) than tumors treated with vehicle (\( n = 7 \); 39.9 ± 7.9%; Fig. 2). This difference was found to be statistically significant (\( p < 0.05 \)).

**DCE-MRI.** Tumor perfusion was monitored in the TLT tumor model 20 minutes after administration of XN via dynamic contrast-enhanced MRI at 4.7 T using i.v. injection of the rapid-clearance blood pool agent P792 (Vistarem\textsuperscript{®}, Guerbet, Roissy, France). High resolution multi-slice \( T_2 \)-weighted spin echo anatomic imaging was performed just before dynamic contrast-enhanced imaging. Pixel-by-pixel values for \( K^{\text{trans}} \), \( V_p \), and \( K_{ep} \) were generated via tracer kinetic modeling of the dynamic contrast-enhanced data, and the resulting parametric maps for \( K^{\text{trans}} \), \( V_p \), and \( K_{ep} \) were generated. Statistical significance for \( V_p \) or \( K^{\text{trans}} \) identified “perfused” pixels, i.e., pixels to which the contrast agent P792 had access.\textsuperscript{6,19}

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Irradiation was performed in the time window during which intratumoral \( pO_2 \) was maximal (i.e., 20 min after XN administration). MRI was performed on a 3 T clinical system (Philips Medical Systems, Best, The Netherlands). DCE-MRI images were obtained 10 to 30 minutes after XN administration. These data determined the time window of vascular bed opening, which was used to design the “flow” experiments as well as as a rationale for scheduling the use of XN as a co-treatment in irradiation and cyclophosphamide chemotherapy experiments.

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clearance blood pool agent P792 (Vistarem). The pixel-by-pixel analysis generated “perfusion maps” and histograms (using the values for Vp, the blood plasma volume per unit volume of tissue) and “permeability maps” and histograms (using the values for Ktrans, the influx volume transfer constant, from plasma into the interstitial space, and Kep, the efflux volume transfer constant from the interstitial space back to plasma). Significant increases in plasmatic volume fraction (Vp) and kep were observed, showing that both flow and permeability parameters were enhanced (Fig. 3). Post-treatment distribution of Vp is broader and more heterogeneous, showing that all tumor regions do not respond to the same extent. However, heterogeneities in the distributions of pharmacokinetic parameters have previously been shown in experimental as well as in human tumors.

**Effect of XN on radiation sensitivity**

To assess the therapeutic relevance of the transient increase in tumor oxygenation after XN administration, we combined XN with 10 Gy radiotherapy. Irradiation was performed 20 min after injection of XN or vehicle. Control experiments included injection of XN alone (without irradiation) and injection of saline alone.

The tumor size was monitored every day for all mice. The time for each tumor to reach 14 mm was calculated. The time differences between treated groups and the control group were calculated (regrowth delays or TGD) (Fig. 4). The results show that tumor growth was significantly delayed by XN: TGD were $5.8 \pm 1.0$ days and $4.2 \pm 0.8$ days for XN and control groups, respectively; resulting in a factor of enhancement in radiation response of 1.4.

**Effect of XN on chemotherapeutic treatment**

To evaluate the possible adjuvant effect of XN on chemotherapy, we carried out a protocol that used a suboptimal dose of cyclophosphamide to facilitate the identification of a possible potentiation of combined treatments. This protocol has already been used to show the benefits of drugs that transiently open the tumor vascular bed. Administration of cyclophosphamide was performed 20 min after injection of XN or vehicle. Control experiments were performed by injecting XN or vehicle alone. The tumor size was measured every day for all mice. The time for each tumor to reach 14 mm was calculated. The time differences between treated groups and the control group were calculated (regrowth delays or TGD) (Fig. 5). These results show that...
XN potentiates the effect of chemotherapy, as tumors co-treated with XN show a TGD of 4.5 ± 1.6 days vs. 1.7 ± 1 days for cyclophosphamide alone, resulting in a factor of enhancement in response to chemotherapy of 2.7.

Discussion

The use of co-treatments is more and more considered in order to improve the therapeutic efficacy of chemotherapy or radiotherapy. Nevertheless, to be really efficient, a combination of treatments has to be carefully scheduled, i.e., radiotherapy and chemotherapy have to be applied at the exact moment when the vascular bed is opened by the co-treatment. This time window can only be identified by the study of key tumor micro-environment parameters, such as oxygenation and perfusion. In this study, we considered the relevance of combining a peripheric vasodilator, Xanthinol Nicotinate, with radio- and chemo-therapy.

We observed a transient increase in tumor pO2 from 10 to 30 minutes after administration of Xanthinol Nicotinate, which was correlated with an increased tumor perfusion. The application of radio or chemotherapy in the time window of the vascular bed opening (i.e., 20 minutes after XN administration) resulted in a significant improvement in radiation sensitivity and response to chemotherapy, by a factor of 1.4 and 2.7, respectively.

The selective aspect of the pro-vascular approach rely on the fact that tumor tissues are more hypoxic and less perfused than normal tissues, therefore, the relative enhancement in oxygen and blood flow (respectively related to radiation and chemotherapeutic responses) is larger in tumors than in healthy tissues. Moreover, the critical cut off value in terms of pO2 and radiosensitization is situated between 5–10 mmHg. Therefore, the impact of a pro-vascular agent able to increase tumor pO2 above this cut off is substantially compared to normal tissues.

XN has thrombolytic and hypotensive activities in rats and anti-platelet and fibrinolytic effects in patients with peripheral arterial oblitative disease. It has been hypothesized that the mechanism of XN partly consists of a simultaneous release of endogenous prostacyclin and nitric oxide by the nicotinate component of the drug.8–10 Our group already characterized several NO-mediated modifiers of tumor hemodynamics and radiation response with success, including NO donors and insulin.8–10 Although further experiments would be required to fully understand the mechanism of action of the drug within a tumor, the proof of concept of the relevance of using this peripheric vasodilator as a co-treatment to radio and chemotherapy is established in experimental tumors. Moreover, the translation to humans would be direct since the drug is already used safely for several human pathologies.

Nevertheless, additional pre-clinical studies on more elaborated tumor models as well as combination to fractionated radiotherapy or other classes of anti-cancer agents will be required to further characterize the potential use of XN as a potential co-treatment in the clinic.

References