

## An improved HPLC-UV method for the simultaneous quantification of triterpenic glycosides and aglycones in leaves of *Centella asiatica* (L.) Urb (APIACEAE)<sup>☆</sup>

M.H. Rafamantanana<sup>a,b,\*</sup>, E. Rozet<sup>c</sup>, G.E. Raelison<sup>a</sup>, K. Cheuk<sup>a</sup>, S.U. Ratsimamanga<sup>a</sup>, Ph. Hubert<sup>c</sup>, J. Quetin-Leclercq<sup>b</sup>

<sup>a</sup> IMRA (Institut Malgache de Recherches Appliquées), BP 3833, 101 Antananarivo, Madagascar

<sup>b</sup> Laboratoire de Pharmacognosie, Unité CHAM, LDRI (Louvain Drug Research Institute), UCL (Université catholique de Louvain), Av. E. Mounier 72, 1200 Bruxelles, Belgium

<sup>c</sup> Laboratoire de Chimie Analytique, Département de Pharmacie, CIRM, ULg (Université de Liège), CHU, B36, B-4000 Liège, Belgium

### ARTICLE INFO

#### Article history:

Received 13 October 2008

Accepted 16 March 2009

Available online 19 March 2009

#### Keywords:

*Centella asiatica*

Madecassoside

Asiaticoside

Madecassic acid

Asiatic acid

Method validation

Quantification

### ABSTRACT

The simultaneous quantification of madecassoside, asiaticoside, madecassic acid and asiatic acid in *Centella asiatica* by HPLC-UV is proposed. Asiaticoside was used as reference for the quantification of heterosides and asiatic acid for aglycones. The evaluation of the extraction efficiency of the four molecules led to use Soxhlet extraction for 8 h. The method was validated and was found to be accurate in the concentration range of 1.0–3.0 mg/ml for asiaticoside and 0.5–2.0 mg/ml for asiatic acid with CV <3% for all investigated compounds. LOD and LOQ were, respectively, 0.0113 and 1.0 mg/ml for asiaticoside and 0.0023 and 0.5 mg/ml for asiatic acid. This method was shown to be convenient for routine analysis of samples of *C. asiatica*.

© 2009 Elsevier B.V. All rights reserved.

### 1. Introduction

*Centella asiatica* (APIACEAE) is an ethnomedicinal herbaceous species, originated from India which grows spontaneously in subtropical regions: China, Malaysia, Australia, America, South Africa and Madagascar. In Madagascar, the plant is largely used by the local population and is the second medicinal species exported [1]. *C. asiatica* is claimed to have a number of medicinal properties and is used in Ayurvedic medicine for the treatment of leprosy, skin tuberculosis, wound healing, stomach aches, arthritis, varicose veins, high blood pressure and as a memory enhancer [2]. Recently, several studies demonstrated that extracts of the plant possess antioxidant activity [3,4], have antiproliferative effects in tumor cells [5], improve venous wall alterations in chronic venous hypertension and protect the venous endothelium [6]. Asiaticoside, one of its active molecules, is reported to cause changes in gene expression and to induce type I collagen synthesis in human fibroblasts [7–9]. Madecassoside was reported to have an anti-rheumatoid effect and wound healing properties [10,11].

<sup>☆</sup> This paper is part of a special issue entitled "Method Validation, Comparison and Transfer", guest edited by Serge Rudaz and Philippe Hubert.

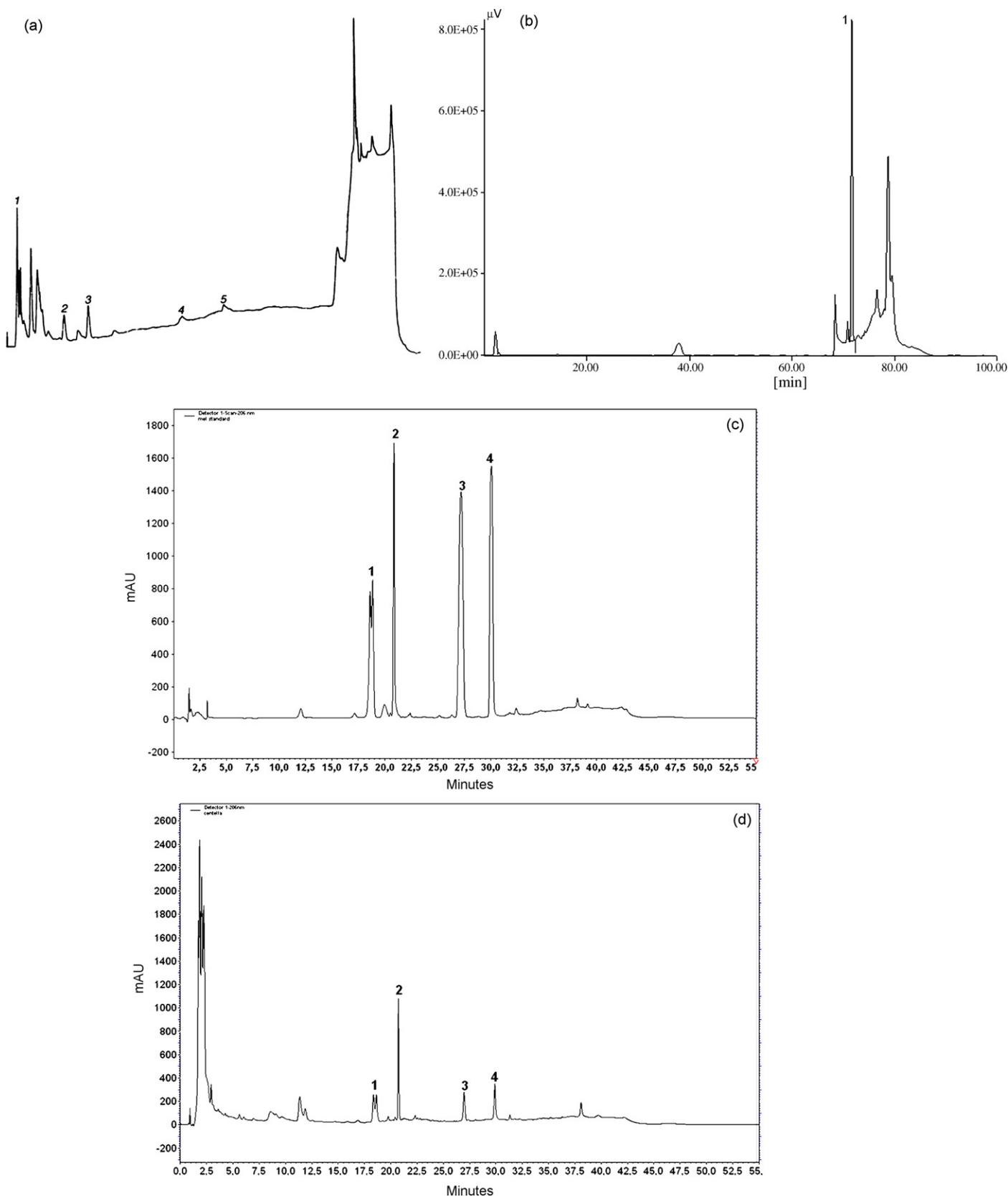
\* Corresponding author at: Laboratoire de Pharmacognosie, Unité CHAM, LDRI (Louvain Drug Research Institute), UCL (Université Catholique de Louvain), Av. E. Mounier 72, 1200 Bruxelles, Belgium.

E-mail address: [mamy.rafamantanana@uclouvain.be](mailto:mamy.rafamantanana@uclouvain.be) (M.H. Rafamantanana).

Triterpenoids of *C. asiatica* (Fig. 1) are components of medicinal drugs and are much used in cosmetic preparations for skin care [12,13]. Collected during all year long, considerable differences of the triterpenoid contents were observed according to geographic regions, phenotype and genotype [14,15], so the assessment of a validated analytical method is necessary. This will also help local population to determine the best cultivating and harvesting conditions.

Most studies led on *C. asiatica* report the quantification of heterosides [16,17], acids [18] or acids and heterosides but with insufficiently validated methods [14,15,19–21]. Several reported methods were based on HPLC differing only in the mobile phase composition or in the detection system. Xingyi et al. [16] proposed a method only dedicated for the quantification of madecassoside and asiaticoside using acetonitrile/water (29/17, v/v) in isocratic mode. Quantification of only madecassoside, asiaticoside and its isomer has been reported by Zhang et al. [17] using ELSD detector. An addition of  $\beta$ -cyclodextrin in the mobile phase was used for the quantification of the sole madecassic acid [18]. The existing methods for the simultaneous quantification of madecassoside, asiaticoside, madecassic acid and asiatic acid used either acetonitrile/water [19], acetonitrile/water with TFA 0.1% [20] or acetonitrile/water each containing 0.05% of  $H_3PO_4$  as mobile phase [21], together with detection at 205 nm. Most methods did not give good resolution or were not suitable for LC-MS and all were insufficiently validated [19–21]. The monography of the European Pharmacopeia [22] quantifies total triterpenoids (Fig. 2a) but our attempts to reproduce the separa-





**Fig. 2.** (a) Chromatogram of a crude extract of *Centella asiatica* [22] (1: solvent, 2: madecassoside (TR = 5.8), 3: asiaticoside (TR = 8.1), 4: madecassic acid (TR = 17.6), 5: asiatic acid (TR = 21.7)); (b) chromatogram of a crude extract of *Centella asiatica* obtained in our lab with European Pharmacopoeia method (1: madecassoside); (c) chromatogram of reference standards with the developed method (1: madecassoside and its isomer (asiaticoside B), 2: asiaticoside, 3: madecassic acid, 4: asiatic acid); (d) chromatogram of a crude extract of *Centella asiatica* with the developed method.

**Table 2**  
Peaks area ratio of the different compounds investigated with asiaticoside.

	Compounds			
	Madecassoside	Asiaticoside	Madecassic acid	Asiatic acid
Reference standards purity	97.94%	99.2%	95%	99%
Peak area ratio <sup>a</sup> (n = 2)	1 ± 0.5 <sup>b</sup>	1.0 ± 0.2 <sup>b</sup>	2.1 ± 1.3 <sup>b</sup>	2.1 ± 0.3 <sup>b</sup>

<sup>a</sup> Area ratio for each compound was calculated using the response of asiaticoside (major compound) as reference.

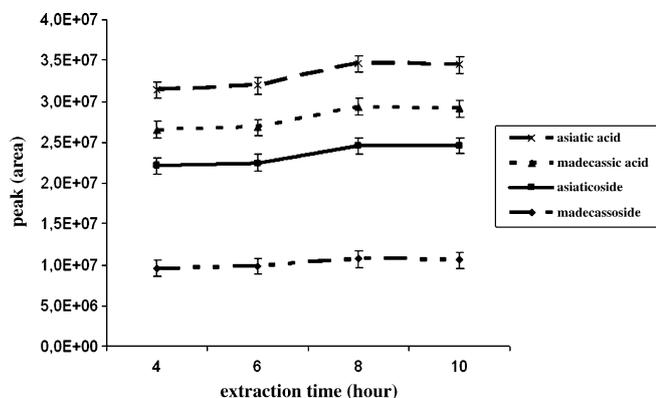
<sup>b</sup> RSD (%).

to the chromatographic conditions and not an extraction failure [22].

### 3.2. Method validation

Selectivity and peak purity were analysed by the comparison of retention times and mass spectra with reference compounds. Mass spectra were analysed at three levels (beginning, middle and end) of each peak investigated and found to be comparable (Figs. 2c–d and 4a–d). Comparison of the chromatogram of an extract of *C. asiatica* from European Pharmacopoeia method (Fig. 2a) and the chromatogram obtained with our method (Fig. 2d) shows its good resolution and the interest of this developed method for the quantification of aglycones. We also observed that madecassoside and its isomer (asiaticoside B also called terminoside) were slightly separated, but as both are considered to be active and are usually not separated; we added both areas and considered both peaks as one madecassoside peak to allow comparison with other results. Calibration standards of asiaticoside and asiatic acid were prepared without matrix ( $m = 3$ ,  $n = 2$ ). Different regression models were tested such as: weighted ( $1/X$ ) quadratic regression, weighted ( $1/X^2$ ) quadratic regression, quadratic regression, weighted ( $1/X$ ) linear regression, weighted ( $1/X^2$ ) linear regression, linear regression after logarithm transformation, linear regression after square root transformation, weighted ( $1/X$ ) linear regression, linear regression. Accuracy profiles were plotted to determine the most suitable regression model [24]. Fig. 5a and b shows the accuracy profiles obtained with the quadratic regression as response function for both standards. It was selected as the most adequate one as the 95%  $\beta$ -expectation tolerance intervals were totally included inside the  $\pm 20\%$  acceptance limits for each concentration level of the validation standards for both analytes.

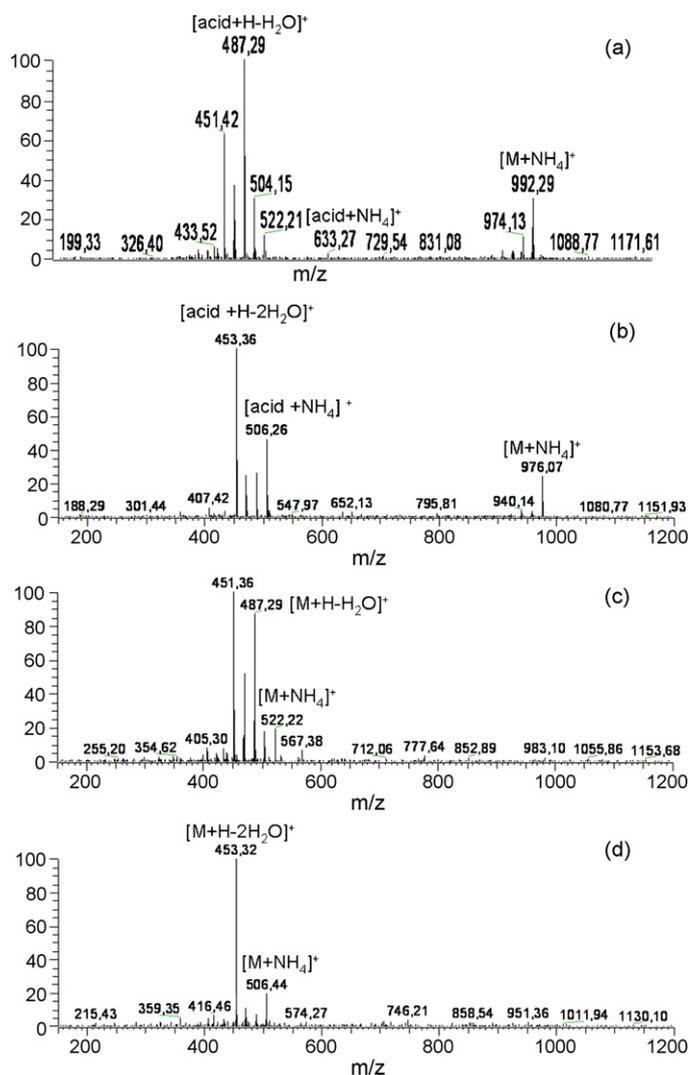
Trueness [25,26] is expressed in relative bias (%) at each concentration level of the validation standards. Relative bias was less than 3% (Table 3) for asiaticoside and 10% for acid asiatic showing the excellent trueness of the method.



**Fig. 3.** Madecassoside, asiaticoside, madecassic and asiatic acids responses after different times of Soxhlet extraction of *Centella asiatica* ( $n = 3$ , RSD < 1%).

Precision was evaluated in terms of standard deviation (SD, mg/ml) and relative standard deviation (RSD %) values for repeatability and intermediate precision [27,28]. As seen in Table 3, RSD (%) for repeatability and intermediate precision did not exceed 4%.

Accuracy allows to evaluate total error, the sum of systematic and random errors of the tests results [24–28]. For asiaticoside and asiatic acid, as illustrated in Fig. 5a and b, their respective accuracy profiles show that the relative upper and lower 95%  $\beta$ -expectation tolerance limits are totally included inside the acceptance limits set at  $\pm 20\%$ . The method can thus be considered as accurate between



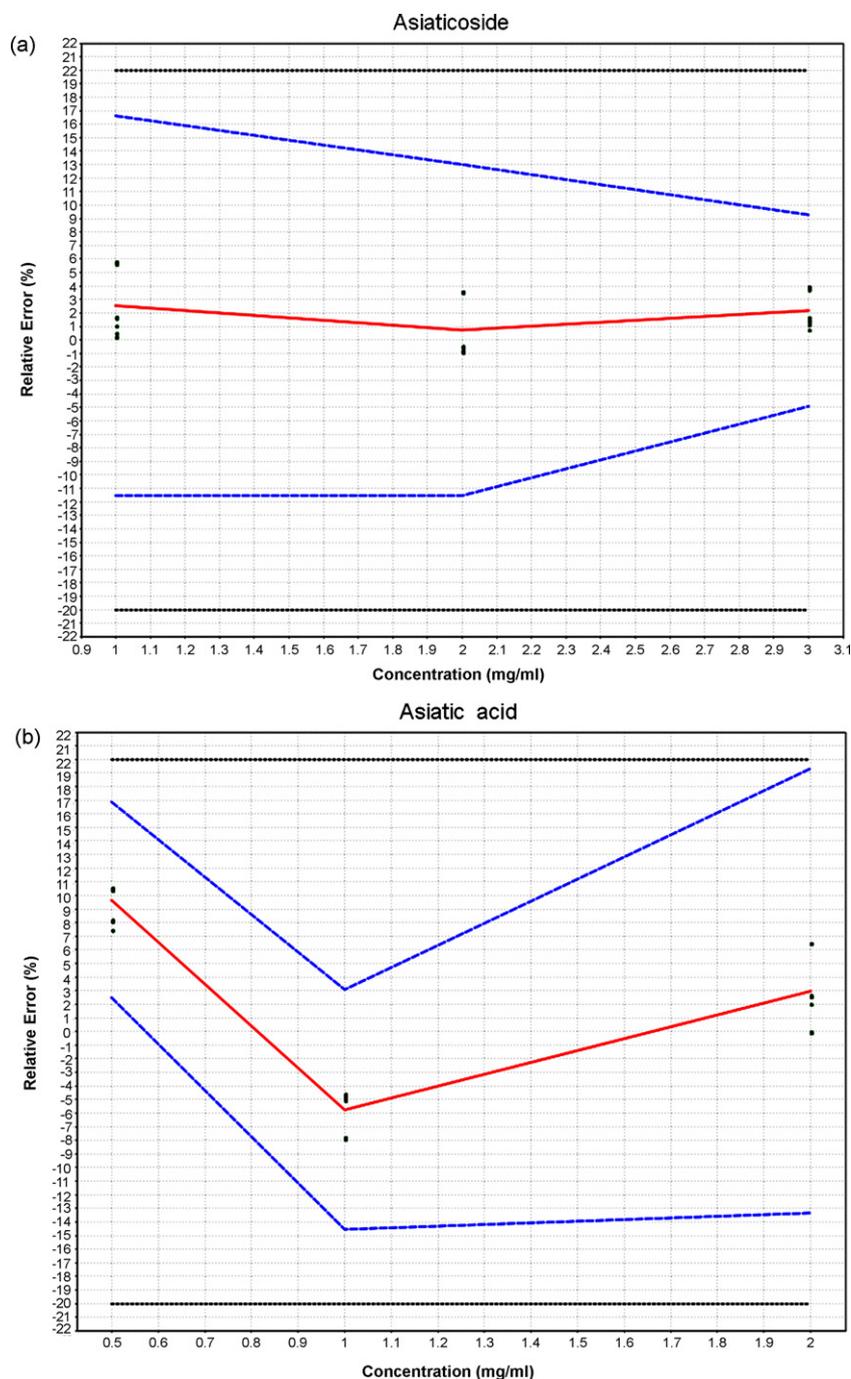
**Fig. 4.** Positive ion mode mass spectra obtained for peaks of the methanol extract of *Centella asiatica*. (a) madecassoside  $[M+NH_4]^+ = 992$ ; (b) asiaticoside  $[M+NH_4]^+ = 976$ ; (c) madecassic acid  $[M+NH_4]^+ = 522$ ; (d) asiatic acid  $[M+NH_4]^+ = 506$ .

1.0 and 3.0 mg/ml for asiaticoside and between 0.5 and 2.0 mg/ml for asiatic acid.

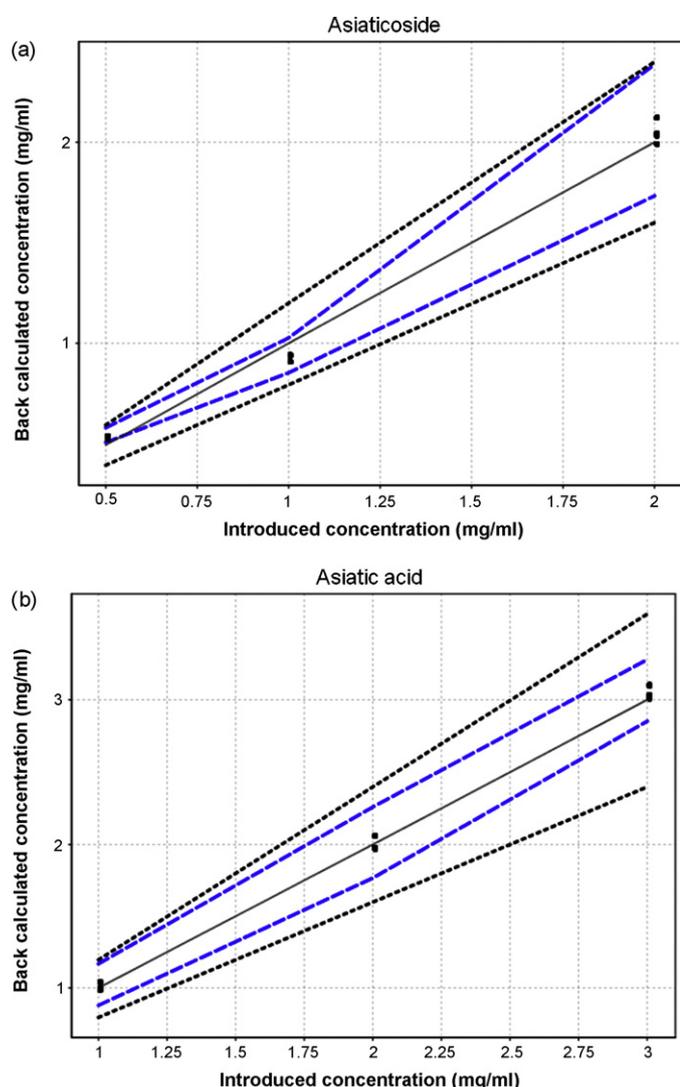
For asiaticoside and asiatic acid, LOD (the smallest quantity of the analyte that can be detectable in the sample, but not quantifiable) were 0.0113 and 0.0023 mg/ml, respectively. These results were estimated using the mean intercept of the calibration model and the residual variance of the regression. The LOQ (the smallest quantity quantifiable in the sample) were determined with the accuracy profiles as they are the smallest concentration levels where the 95%  $\beta$ -expectation tolerance limits remain inside the  $\pm 20\%$  acceptance limits [24–26]. In other words, they are the smallest concentration levels with a maximum

total error of 20%. As shown in Fig. 5a and b, the LOQ are the smallest concentration levels of the validation standards, i.e. 1.0 and 0.5 mg/ml for asiaticoside and asiatic acid, respectively.

Uncertainty of the measurement characterises the dispersion of the values that could reasonably be attributed to the measurer [29,30]. It was evaluated using expanded uncertainty where true value can be observed with a confidence level at 95%. Table 4 shows that relative expanded uncertainties were less than 10% for asiaticoside and asiatic acid which means that the unknown true value is located at a maximum of  $\pm 10\%$  around the measured result.



**Fig. 5.** Accuracy profiles of asiaticoside (a) and asiatic acid (b) obtained with quadratic regression. The plain line is the relative bias, dashed lines are the  $\beta$ -expectation tolerance limit ( $\beta=95\%$ ) and dotted lines represent the acceptance limit ( $\pm 20\%$ ). The dots represent the relative back-calculated concentrations of the validation standards and are plotted according to their targeted concentration.



**Fig. 6.** Linear profiles of asiaticoside (a) and asiatic acid (b). The continuous line is identity line ( $y=x$ ), the dotted lines are the upper and lower acceptance limits in absolute values and the dashed lines are the upper and lower  $\beta$ -expectation tolerance limits ( $\beta=95\%$ ).

The linearity demonstrated the relationship between introduced and calculated concentration [25,26,28] using  $\beta$ -expectation tolerance interval approach. The concentrations of the validation standards were back-calculated in order to determine, by concentration level, the mean relative bias as well as the upper and lower  $\beta$ -expectation tolerance intervals. The acceptance limits were set at  $\pm 20\%$ . In order to demonstrate method linearity, a regression line was fitted on the calculated concentrations of the validation standards as a function of the introduced concentrations by applying a linear regression model. The equations obtained for asiaticoside and asiatic acid with their coefficient of determination are presented in Table 3.

The slopes values obtained for the two standards were, respectively 1.02, 1.023. Fig. 6a and b demonstrate the linearity of the results.

### 3.3. Application to samples of *Centella asiatica*

Three samples of *C. asiatica* were collected in the East and High Plateau regions of Madagascar. Extracts were analyzed and the results obtained by our method and the data described in the literature [14,15] are given in Table 5. Our results are in the same range than those obtained in the previous works. The heterosides are more abundant than aglycones and asiaticoside is often the major compound. Nevertheless, differences occur between samples and those analyzed here seemed less rich than the others, but we have to point out that all these other methods were not validated so the reliability of these results needs caution. This proposed method gives low RSD (%) values and is reliable. These results stress the importance of a good quantification method to determine the best culture and harvesting conditions.

A validated method for the quantification of asiaticoside, madecassoside, asiatic and madecassic acids in drug samples of *C. asiatica* was developed. This method allows the simultaneous quantification of madecassoside, asiaticoside, madecassic acid and asiatic acid. Most of other validated methods only quantified some of these active molecules. Furthermore, the method described in European Pharmacopoeia does not give good signal to noise ratio for the accurate quantification of aglycones and was not reproducible in our laboratory. The method developed and validated was then successfully applied to quantify these four compounds in different samples of *C. asiatica* collected in Madagascar.

**Table 3**  
Validation results in crude extract of *Centella asiatica*.

Validation criteria	Asiaticoside	Asiatic acid
Response function	Quadratic regression Calibration range (3 points) 0.5–5 mg/ml	Quadratic regression Calibration range (3 points) 0.25–2.5 mg/ml
Trueness	Concentration (mg/ml)	Concentration (mg/ml)
	Relative bias (%)	Relative bias (%)
	1	0.51
	2	1
	3	2
Precision	Repeatability (SD mg/ml)	Repeatability (SD mg/ml)
	0.2313	0.2421
	0.0987	0.1427
	0.252	0.1997
Accuracy	$\beta$ -Expectation lower and upper tolerance limits of the relative error (%)	$\beta$ -Expectation lower and upper tolerance limits of the relative error (%)
	–11.5, 16.6	2.5, 16.9
	–11.6, 13.0	–14.6, 3.1
	–5.0, 9.3	–13.4, 19.3
Linearity		
Slope	1.02	1.023
Intercept	–0.005007	–0.01017
$r^2$	0.998	0.9912

**Table 4**  
Estimates of the measurement uncertainties related to asiaticoside and asiatic acid, at each concentration level investigated in validation using quadratic regression model.

Asiaticoside	Concentration level (mg/ml)	Mean introduced concentration (mg/ml)	Uncertainty of the bias (mg/ml)	Uncertainty (mg/ml)	Expanded Uncertainty (mg/ml)	Relative expanded uncertainty (%)
	1.0	1.000	0.01644	0.03294	0.06588	6.6
	2.0	2.000	0.02860	0.05723	0.1145	5.7
	3.0	3.000	0.02554	0.05146	0.1029	3.4
Asiatic acid	Concentration level (mg/ml)	Mean introduced concentration (mg/ml)	Uncertainty of the bias (mg/ml)	Uncertainty (mg/ml)	Expanded uncertainty (mg/ml)	Relative expanded uncertainty (%)
	0.5	0.5000	0.004286	0.008628	0.01726	3.4
	1.0	1.000	0.01031	0.02065	0.04131	4.1
	2.0	2.000	0.03809	0.07625	0.1525	7.6

**Table 5**  
Comparison of the content of triterpenes in *Centella asiatica* samples analysed by our method ( $n = 3$ ) in % of the dry plant with results given in previous works [14,15].

Sample	Madecassoside	Asiaticoside	Madecassic acid	Asiatic acid
Sample 1	1.7 ± 0.04	2.0 ± 0.02	0.95 ± 0.039	0.98 ± 0.03
Sample 2	1.27 ± 0.013	1.63 ± 0.04	<LOD	<LOD
Sample 3	1.64 ± 0.02	1.75 ± 0.02	0.72 ± 0.02	0.72 ± 0.08
CA-1 [14]	4.76 ± 1.342	5.23 ± 0.025	1.97 ± 0.007	1.89 ± 0.08
CA-2 [14]	4.28 ± 0.124	4.52 ± 0.138	1.88 ± 0.07	1.79 ± 0.102
CA-3 [14]	5.89 ± 0.15	6.42 ± 0.48	0.23 ± 0.00	0.16 ± 0.07
CA-4 [14]	3.23 ± 0.06	3.37 ± 0.26	0.37 ± 0.09	0.44 ± 0.17
CA-5 [14]	4.74 ± 0.23	4.58 ± 0.63	0.23 ± 0.08	0.25 ± 0.10
CA-6 [14]	2.38 ± 0.18	2.67 ± 0.27	0.42 ± 0.29	0.36 ± 0.28
CA-7 [14]	2.76 ± 0.29	3.09 ± 0.24	0.12 ± 0.05	0.09 ± 0.07
Leaves type-1 [15]	3.62 ± 0.29	3.68 ± 0.04	0.11 ± 0.01	0.05 ± 0.02
Leaves type-2 [15]	2.76 ± 0.15	3.13 ± 0.14	0.59 ± 0.12	0.64 ± 0.14

Mean ± RSD %. CA: *Centella asiatica* [14].

## Acknowledgements

We wish to thank the “Commission Universitaire pour le Développement” and the Catholic University of Belgium for financial support. We are grateful to Denis Randriamampionona, Dr Gabrielle Chataigné, Marie Christine Fayt, Jean Paul Vanhelputte and Ramazan Colak for technical helps and assistance. A research grant from the Belgium National Fund for Scientific Research (F.R.S.-FNRS) to E. Rozet is also gratefully acknowledged.

## References

- [1] G. Péchard, M. Antona, S. Aubert, D. Babin, Bois et Forêts des Tropiques 284 (2005) 2.
- [2] P. Boiteau, A.R. Ratsimamanga, Thérapie 1 (1956) 11.
- [3] G. Jayashree, G.K. Muraleedhara, S. Sudarslal, V.B. Jacob, Fitoterapia 74 (2003) 5.
- [4] A. Gnanapragasam, K. Kuma Ebenezar, V. Sathish, P. Govindaraju, T. Devaki, Life Sci. 76 (2004) 5.
- [5] M. Yoshida, M. Fuchigami, T. Nagao, H. Okabe, K. Matsunaga, J. Takata, et al., Biol. Pharm. Bull. 28 (2005) 1.
- [6] L. Incadela, M.R. Cesarone, M. Cacchio, M.T. De Sanctis, C. Santavenere, M.G. D'Auro, M. Bucci, G. Belcaro, Angiology 52 (Suppl. 2) (2001) S9.
- [7] C.D. Coldren, P. Hashim, J.M. Ali, S.K. Oh, A.J. Sinskey, C. Rha, Planta Med. 69 (2003) 8.
- [8] L. Lu, K. Ying, S. Wei, Y. Liu, H. Lin, Y. Mao, Br. J. Dermatol. 151 (2004) 3.
- [9] J. Lee, E. Jung, Y. Kim, J. Park, S. Hong, J. Kim, C. Hyun, S.Y. Kim, D. Park, Planta Med. 72 (2006) 4.
- [10] M. Liu, Y. Dai, X. Yao, Y. Li, Y. Xia, Z. Gong, Int. Immunopharmacol. 8 (2008) 11.
- [11] M. Lui, Y. Dai, Y. Li, F. Huang, Z. Gong, Q. Meng, Planta Med. 74 (2008) 8.
- [12] E.K. Park, S.H. Park, K.R. Patent No. 007051377. Rep. Kor. Kongkae Taeho Kongbo (2007).
- [13] D.H. Park, J.S. Lee, G.S. Jung, K.R. Patent No. 2007111785 Rep Kor. Kongkae Taeho Kongbo (2007).
- [14] D. Randriamampionona, B. Diallo, F. Rakotonirina, C. Rabemanantsoa, K. Cheuk, A.M. Corbisier, J. Mahillon, S. Ratsimamanga, M. El Jaziri, Fitoterapia 78 (2007) 7.
- [15] T.J. Jacinda, R. Meyer, I.A. Dubery, Plant Cell. Tissue Organ Culture 94 (2008) 1.
- [16] L. Xingyi, F. Lijuan, Zhongguo Yaoshi (Wuhan, China) 11 (2008) 11.
- [17] F.L. Zhang, Y.J. Wei, J. Zhu, Z.N. Gong, Biomed. Chromatogr. 22 (2008) 2.
- [18] P. Jian, K. Guiqing, Y. Chuanxun, Z. Beibei, J. Risheng, Y. Yuan, J. Chromatogr. 25 (2007) 3.
- [19] B.T. Schaneberg, J.R. Mikell, I.A. Khan, Pharmazie 58 (2003) 6.
- [20] P.K. Inamdar, R.D. Yeole, A.B. Ghogare, N. de Souza, J. chromatogr. A 742 (1996) 1.
- [21] B. Günther, H. Wagner, Phytomedicine 3 (1996) 59.
- [22] European Pharmacopoeia, 6th ed., 2008 (<http://www.edqm.eu>).
- [23] Analytical Methods committee, Analyst 120 (1995) 2303.
- [24] E. Rozet, C. Hubert, A. Ceccato, W. Dewé, E. Ziemons, F. Moonen, K. Michail, R. Wintersteiger, B. Streeel, B. Boulanger, Ph. Hubert, J. Chromatogr. A 1158 (2007) 126.
- [25] E. Rozet, A. Ceccato, C. Hubert, E. Ziemons, R. Oprean, S. Rudaz, B. Boulanger, Ph. Hubert, J. Chromatogr. A 1158 (2007) 111.
- [26] Ph. Hubert, J.J. Nguyen-Huu, B. Boulanger, E. Chapuzet, P. Chiap, N. Cohen, P.A. Compagnon, W. Dewe, M. Feinberg, M. Lallier, M. Laurentie, N. Mercier, G.M. Lizard, C. Nivet, L. valet, J. Pharm. Biomed. Anal. 36 (2004) 579.
- [27] Guidance for industry: Bioanalytical Method Validation, US Department of Health and Human Services, US Food and Drug Administration (FDA), Center for Drug Evaluation and Research (CDER), Center for Biologics Evaluation and Research (CBER), Rockville, May 2001.
- [28] Ph. Hubert, J.J. Nguyen-Huu, B. Boulanger, E. Chapuzet, P. Chiap, N. Cohen, P.A. Compagnon, W. Dewe, M. Feinberg, M. Lallier, M. Laurentie, N. Mercier, G.M. Lizard, L. Valat, E. Rozet, J. Pharm. Biomed. Anal. 45 (2007) 82.
- [29] Eurachem/Citac Guide Quantifying the uncertainty in analytical measurement, 2nd ed., 2000.
- [30] CBEA-4/16, EA Guidelines on the Expression of Uncertainty in Quantitative Testing, 2004, <http://www.european-accreditation.org>.